Protocol for cleaning the iTC200 and leaving it for the next user

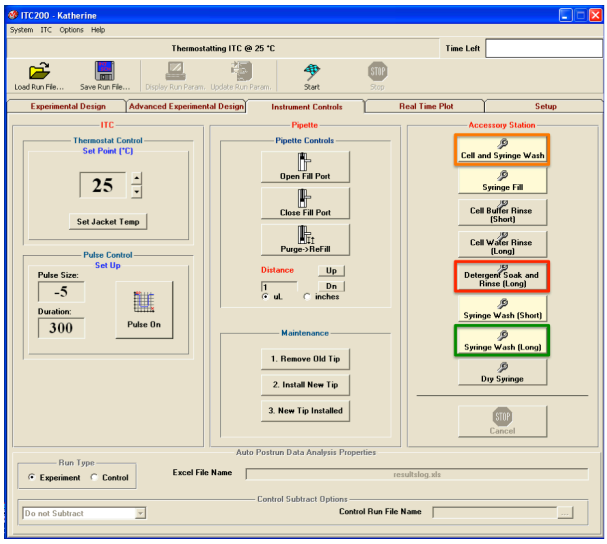
1) When you have finished your experiments, empty the cell and fill it with 5% Decon[1]. Wait for one minute and then empty the cell again.

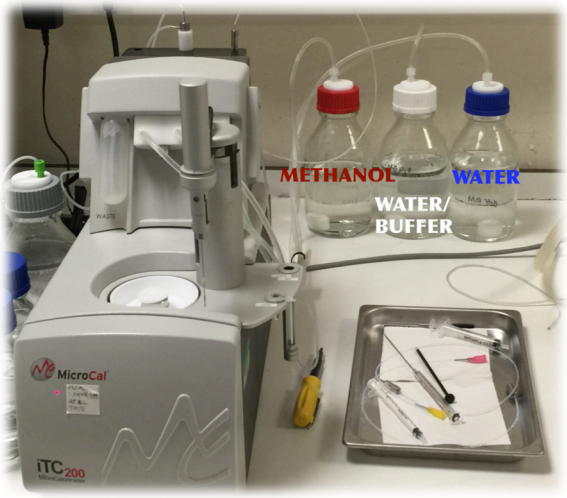
2) Run "**Cell and Syringe Wash**", following the stepwise instructions[2] ➔

OR…

1) If there has been any sign of precipitation or aggregation, or a low DP/drifting baseline, run "**Detergent Soak and Rinse**", which starts with a 30 minute 5% Decon soak at 50 °C. ➔

2) Follow with "**Syringe Wash (Long)**". ➚



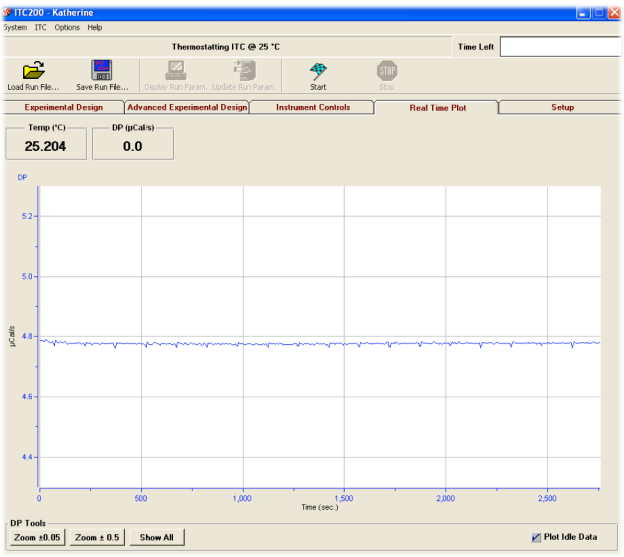


➔ *Always check that the **water** and **methanol** reservoirs contain sufficient volumes beforehand[3].*

3) Fill the cell and syringe with fresh MilliQ water (filling the syringe 2× or 3× removes all traces of methanol and gives the best results).

4) Load the run file Katherine/water_USEME.inj. Please name the data file **YYYYMMDDwater.itc**, and save it in your data directory. (These files are a very useful resource for monitoring instrument performance over time.)

5) Start the run; the DP should be > 4.5 and the injection heats small. ➔



5) Clean up the bench area and empty the waste reservoir. Inform Simon Quick (sq204@cam.ac.uk) of any consumable stocks that are running low.

6) After the run (or the following morning), place the pipette in the rest position and put the cap on the cell.

Many thanks,

Katherine and Simon (15th Oct 2020)

[1]There is a stock of 100% Decon under the sink if the supply of 5% is running low.
[2]If working with fragments of limited solubility, consider filling the syringe first with 100% DMSO to dissolve any remaining traces that could precipitate during the water wash step.
[3]Methanol is available in the cupboard under the fumehood.